

Lot No.: **42F**

Lot-specific information

## **Olerup SSP<sup>®</sup> DQB1\*05**

Product number:	101.211-24u – without <i>Taq</i> polymerase
Lot number:	42F
Expiry date:	2011-January-01
Number of tests:	24
Number of wells per test:	6
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

**This Product Description is only valid for Lot No. 42F**

### **CHANGES COMPARED TO THE PREVIOUS *OLERUP SSP<sup>®</sup> DQB1\*05* LOT**

The DQB1\*05 primer set, specificity and interpretation tables are unchanged compared to the previous *Olerup SSP<sup>®</sup> DQB1\*05* lot (**Lot No. 09E**).

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## PRODUCT DESCRIPTION

### DQB1\*05 SSP subtyping

#### CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1\*0501 to DQB1\*0505 alleles.

*Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.*

#### PLATE LAYOUT

Each test consists of 6 PCR reactions in an 8 well PCR plate. Wells 7 and 8 are empty.

1	2	3	4	5	6	empty	empty
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The 8 well cut PCR plate is marked with 'DQ5' in silver gray ink.

Well No. 1 is marked with the Lot No. '42F'.

The PCR plates are covered with a PCR-compatible foil.

**Please note:** When removing each 8 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

#### INTERPRETATION

Only the DQB1\*05 alleles will be amplified by the DQB1\*05 subtyping kit. Thus, the interpretation of DQB1\*05 subtypings is not influenced by other groups of DQB1 alleles or the DQB2 and DQB3 genes.

#### UNIQUELY IDENTIFIED ALLELES

All the DQB1\*05 alleles, i.e. **DQB1\*0501 to DQB1\*0505**, recognized by the HLA Nomenclature Committee in January 2009<sup>1</sup> will give rise to unique amplification patterns by the primers in the DQB1\*05 subtyping kit.

The DQB1\*05 subtyping kit cannot distinguish the DQB1\*050101 and DQB1\*050102 alleles and the DQB1\*050301 and DQB1\*050302 alleles.

<sup>1</sup>DQB1 alleles listed on the IMGT/HLA web page 2009-January-16, release 2.24.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

#### RESOLUTION IN HOMO- AND HETEROZYGOTES

The 5 DQB1\*05 alleles generate 6 different amplification patterns, as the DQB1\*050201 and DQB1\*050202 alleles generate different amplification patterns. These can be combined in 21 homozygous and heterozygous combinations. Eight of these genotypes do not give rise to unique amplification patterns.

+-+--+      050202,0505 = 0503,0505  
+-+---      050201,050202 = 050201,0503 = 050202,050202 =  
                 050202,0503  
+-+---+      050201,0505 = 0505,0505

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## SPECIFICITY TABLE

### DQB1\*05 SSP subtyping

Specificities and sizes of the PCR products of the 6 primer mixes used for DQB1\*05 SSP subtyping

Primer Mix	Size of spec. PCR product <sup>1</sup>	Size of control band <sup>2</sup>	Amplified DQB1*05 <sup>3</sup> alleles
1	225 bp	515 bp	050101-0505
2	135 bp	430 bp	050101-050102
3	120 bp	430 bp	050201-050202, 0505
4	95 bp	515 bp	050202, 050301-050302
5	120 bp	430 bp	0504
6	185 bp	430 bp	0505

<sup>1</sup>Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1\*05 SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective length of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

<sup>2</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1\*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

<sup>3</sup>For several DQB alleles only partial second exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of codons 87 to 92 are conserved within allelic groups.

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<b>INTERPRETATION TABLE</b>						
<b>DQB1*05 SSP subtyping</b>						
<b>Amplification patterns of the DQB1*05 alleles</b>						
	<b>Well</b>					
	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>
<b>Length of spec.</b>	<b>225</b>	<b>135</b>	<b>120</b>	<b>95</b>	<b>120</b>	<b>185</b>
<b>PCR product</b>						
<b>Length of int.</b>	<b>515</b>	<b>430</b>	<b>430</b>	<b>515</b>	<b>430</b>	<b>430</b>
<b>pos. control<sup>1</sup></b>						
<b>5'-primer<sup>2</sup></b>	<b>26</b>	<b>26</b>	<b>30</b>	<b>30</b>	<b>30</b>	<b>38</b>
	5'-g gg <sup>3'</sup>	5'-g gg <sup>3'</sup>	5'-gA C <sup>3'</sup>	5'-gA C <sup>3'</sup>	5'-gA T <sup>3'</sup>	5'-C gC <sup>3'</sup>
<b>3'-primer<sup>3</sup></b>	<b>87</b>	<b>57</b>	<b>57</b>	<b>47</b>	<b>57</b>	<b>86</b>
	5'-g gT <sup>3'</sup>	5'-C AA <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-Cg A <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-A Cg <sup>3'</sup>
<b>Well No.</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>
<b>DQB1 allele</b>						
<b>*050101-050102</b>	<b>1</b>	<b>2</b>				
<b>*050201</b>	<b>1</b>		<b>3</b>			
<b>*050202</b>	<b>1</b>		<b>3</b>	<b>4</b>		
<b>*050301-050302</b>	<b>1</b>			<b>4</b>		
<b>*0504</b>	<b>1</b>				<b>5</b>	
<b>*0505</b>	<b>1</b>		<b>3</b>			<b>6</b>
<b>DQB1 allele</b>						
<b>Well No.</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>

<sup>1</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1\*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

<sup>2</sup>The codon, in the 2<sup>nd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given. Empty spaces indicate codon boundaries.

<sup>3</sup>The codon, in the 2<sup>nd</sup> exon, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given. Empty spaces indicate codon boundaries.

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CELL LINE VALIDATION SHEET										
DQB1*05 SSP subtyping kit										
				Production No.	Well					
					1	2	3	4	5	6
					200627301	200627302	200627303	200627304	200854505	200627306
	IHWC cell line		DQB1							
1	9001	SA	*0501		+	+	-	-	-	-
2	9280	LK707	*0601	*0202	-	-	-	-	-	-
3	9011	E4181324	*0601		-	-	-	-	-	-
4	9275	GU373	*0201		-	-	-	-	-	-
5	9009	KAS011	*0502		+	-	+	-	-	-
6	9353	SM	*0302	*0601	-	-	-	-	-	-
7	9020	QBL	*0201		-	-	-	-	-	-
8	9007	DEM	*0302	*0502	+	-	+	-	-	-
9	9026	YAR	*0302		-	-	-	-	-	-
10	9107	LKT3	*0401		-	-	-	-	-	-
11	9051	PITOUT	*0202		-	-	-	-	-	-
12	9052	DBB	*0303		-	-	-	-	-	-
13	9067	BTB	*0402		-	-	-	-	-	-
14	9071	OLGA	*0402		-	-	-	-	-	-
15	9075	DKB	*0303		-	-	-	-	-	-
16	9037	SWEIG007	*0301		-	-	-	-	-	-
17	9008	WILJON	*0602	*0603	-	-	-	-	-	-
18	9257	32367	*0602	*0202	-	-	-	-	-	-
19	9038	BM16	*0301		-	-	-	-	-	-
20	9059	SLE005	*0604		-	-	-	-	-	-
21	9064	AMALA	*0301		-	-	-	-	-	-
22	9056	KOSE	*0503	*0604	+	-	-	+	-	-
23	9124	IHL	*0503	*0601	+	-	-	+	-	-
24	9035	JBUSH	*0301		-	-	-	-	-	-
25	9049	IBW9	*0202		-	-	-	-	-	-
26	9285	WT49	*0201		-	-	-	-	-	-
27	9191	CH1007	*0401	*0501	+	+	-	-	-	-
28	9320	BEL5GB	*0202	*0301	-	-	-	-	-	-
29	9050	MOU	*0202		-	-	-	-	-	-
30	9021	RSH	*0402		-	-	-	-	-	-
31	9019	DUCAF	*0201		-	-	-	-	-	-
32	9297	HAG	*0301		-	-	-	-	-	-
33	9098	MT14B	*0302		-	-	-	-	-	-
34	9104	DHIF	*0301		-	-	-	-	-	-
35	9302	SSTO	*0305		-	-	-	-	-	-
36	9024	KT17	*0302		-	-	-	-	-	-
37	9065	HHKB	*0603		-	-	-	-	-	-
38	9099	LZL	*0301		-	-	-	-	-	-
39	9315	CML	*0201	*0301	-	-	-	-	-	-
40	9134	WHONP199	*0202	*0303	-	-	-	-	-	-
41	9055	H0301	*0609		-	-	-	-	-	-
42	9066	TAB089	*0601		-	-	-	-	-	-
43	9076	T7526	*0303		-	-	-	-	-	-
44	9057	TEM	*0503		+	-	-	+	-	-
45	9239	SHJO	*0202		-	-	-	-	-	-
46	9013	SCHU	*0602		-	-	-	-	-	-
47	9045	TUBO	*0301		-	-	-	-	-	-
48	9303	TER-ND	*0501		+	+	-	-	-	-



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## CERTIFICATE OF ANALYSIS

### **Olerup SSP<sup>®</sup> DQB1\*05 SSP**

**Product number:** 101.211-24u – without *Taq* polymerase  
**Lot number:** 42F  
**Expiry date:** 2011-January-01  
**Number of tests:** 24  
**Number of Wells per test:** 6

#### **Well specifications:**

Well No.	Production No.
1	2006-273-01
2	2006-273-02
3	2006-273-03
4	2006-273-04
5	2008-545-05
6	2006-273-06

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 5 and 6 were available. The specificities of the primers in these primer solutions were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer.

**Results:** No false positive or false negative amplifications were obtained.

**Date of approval:** 2009-May-25

**Approved by:**

**Quality Control, Supervisor**

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## Declaration of Conformity

**Product name:** *Olerup* SSP® DQB1\*05  
**Product number:** 101.211-24u  
**Lot number:** 42F

**Intended use:** DQB1\*05 resolution histocompatibility testing

**Manufacturer:** *Olerup* SSP AB  
Hasselstigen 1  
SE-133 33 Saltsjöbaden, Sweden  
**Phone:** +46-8-717 88 27  
**Fax:** +46-8-717 88 18

We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2000 and ISO 13485:2006, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden  
2009-May-25

Olle Olerup  
Managing Director



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**ADDRESSES:**

**Manufacturer:**

**Olerup SSP AB**, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

**Tel:** +46-8-717 88 27

**Fax:** +46-8-717 88 18

**E-mail:** [info-ssp@olerup.com](mailto:info-ssp@olerup.com)

**Web page:** <http://www.olerup.com>

**Distributed by:**

**Olerup GmbH**, Löwengasse 47 / 6, AT-1030 Vienna, Austria.

**Tel:** +43-1-710 15 00

**Fax:** +43-1-710 15 00 10

**E-mail:** [support-at@olerup.com](mailto:support-at@olerup.com)

**Web page:** <http://www.olerup.com>

**Olerup Inc.**, 901 S. Bolmar St., Suite R, West Chester, PA 19382

**Tel:** 1-877-OLERUP1

**Fax:** 610-344-7989

**E-mail:** [info.us@olerup.com](mailto:info.us@olerup.com)

**Web page:** <http://www.olerup.com>

For information on *Olerup* SSP distributors worldwide, contact **Olerup GmbH**.